

Product Number: DF37293



DNA-Fect™ 293
in vitro DNA Transfection Reagent

Instruction Manual

Neuromics
5325 West 74th Street, Suite 8
Edina, MN 55438
Phone: 952-374-6161
Fax: 612-677-3976
Email: pshuster@neuromics.com
Website: www.neuromics.com

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Pete Shuster
 Neuromics
 5325 West 74th Street, Suite 8
 Edina, MN 55438
 Email: phuster@neuromics.com

Introduction

in vitro DNA-Fect293 Transfection Reagent contains a patent-pending blend of new gene delivery compounds, which greatly facilitate which greatly facilitate transfection of DNA to HEK293 cell lines such as HEK293T, HEK293FT, HEK293E, HEK293A.

Important Guidelines

- In order to achieve higher efficiency, transfect cells at high density 90-95% confluency is recommended.
- To minimize cytotoxicity, transfect cells in presence of serum (10%) and antibiotics.
- Change medium with serum (10%) and antibiotics 5 hours post transfection is optional.
- *Storage:* DNA-Fect293 *In Vitro* Transfection Reagent is stable for up to 12 months at +4°C. This product is shipped at ambient temperature.

Procedures for Transfecting Mammalian Cells:

1. For Adherent Cells

Cell Seeding

Cells should be plated 18 to 24 hours prior to transfection so that the monolayer cell density reaches 90-95% confluency at the time of transfection. Freshly complete culture medium with serum and antibiotics is added to each well 30-60 minutes before transfection.

Note: High serum levels (>5%) with antibiotics usually do not have inhibitory effect on transfection efficiency. For some cell lines, higher transfection efficiencies are observed in the presence of serum and antibiotics. We recommend you use complete medium containing serum and antibiotics initially.

Preparation of DNA-Fect293-DNA Complex and Transfection Procedures

For different cell types, the optimal ratio of DNA-Fect293 (μL):DNA (μg) varies from 1:1 to 3:1. We recommend the DNA-Fect293 (μL):DNA (μg) ratio of 3:1 as a starting point which usually gives satisfactory transfection efficiency with little cytotoxicity. To ensure the optimal size of complex particles, we recommend using serum-free DMEM with high glucose to dilute DNA and Transfection Reagent.

The following protocol is for transfection in 24-well plates. Refer to Table 1 for transfection in other culture formats. The optimal transfection conditions for a majority of adherent cell lines are given in the standard protocol described below.

Culture Format	Transfection Volume (ml)	Plasmid DNA (μg)	Diluent Volum e (μL)	DNA-Fect293 (μL)
96-well	0.2	0.2	2x10	0.6
48-well	0.3	0.5	2x20	1.0
24-well	0.5	1.0	2x50	3.0
12-well	1.0	0.75	2x75	2.5
6-well	1.0	2.0	2x100	6.0
35 mm	1.0	2.0	2x100	6.0
60 mm	2.8	5.0	2x250	15.0
100 mm	6.0	7.0-8.0	2x500	21.0-24.0
T75 flask	8.0	18.0-36.0	2x750	54.0-108.0

Table 1. Recommended Amounts for Different Culture Formats.

24-well Transfection Protocol

- Add 1 μg of DNA into 50 μl of serum-free DMEM with high glucose. Vortex gently and spin briefly to bring drops to bottom of the tube.

- Immediately add the diluted DNA-Fect293 Reagent to the diluted DNA solution. (Important: do not mix the solutions in the reverse order !).
- Vortex the solution immediately. Spin down briefly to bring liquid drops to bottom of the tube.
- Leave the solution undisturbed for 15-20 min. at room temperature to allow DNA-Fect293-DNA complexes to form. Note: *Never keep the DNA-Fect293-DNA complex longer than 20 minutes.*
- Add 100 μl DNA-Fect293-DNA complex drop-wise into each well containing cells and medium. Mix gently by rocking the plate back and forth.
- Change medium 12-18 hours post transfection.

For sensitive cells, to lower cytotoxicity, remove DNA-Fect293-DNA complex and replace with complete medium 5 hours after transfection.

2. For Suspension Cells

The following protocol is given for transfection in 6-well plate. The protocol can be scaled up or down according to culture volume.

Cell Seeding

Suspension cells are typically seeded the day of the transfection at a density of $0.5-1.0 \times 10^6$ cells per ml of culture. For optimal transfection conditions, seed the number of cells adapted to the culture vessel format according to Table 2.

- 30-60 minutes before transfection, warm fresh medium in a 37°C water bath. Aspirate out the old medium from each well and add 0.5 ml fresh medium with serum and antibiotics.

Culture Format	Number of Cells to Seed
100 mm	$5 \times 10^6 - 1 \times 10^7$
60 mm	$2 \times 10^6 - 5 \times 10^6$
35 mm	$5 \times 10^5 - 2 \times 10^6$
6-well	$2 \times 10^5 - 5 \times 10^5$
24-well	$1 \times 10^5 - 2 \times 10^5$
48-well	$5 \times 10^4 - 1 \times 10^5$
96-well	$2 \times 10^4 - 5 \times 10^4$

Table 2: Recommended Number of Suspension Cells to Seed.

DNA-Fect293-DNA Complex Preparation and Transfection Procedure

For different cell types, the optimal ratio of

DNA-Fect293 (μL):DNA (μg) varies from 1:1 to 3:1. We recommend the DNA-Fect293 (μL):DNA (μg) ratio of 3:1 as a starting point which usually gives satisfactory transfection efficiency with little cytotoxicity. To ensure the optimal size of complex particles, we recommend using serum-free DMEM with high glucose to dilute DNA and Transfection Reagent.

6-well Transfection Protocol

- Add 2 μg of DNA into 100 μl of serum-free DMEM with high glucose. Vortex gently and spin briefly to bring drops to bottom of the tube.
- For each well, dilute 6 μl of DNA-Fect293 Reagent into 100 μl of DMEM Serum-free Medium with High Glucose. Vortex gently and spin down briefly.
- Add 100 μl of DNA-Fect293 immediately to the 100 μl DNA solution all at once (Important: do not mix the solutions in the reverse order !).
- Vortex the solution immediately. Spin down briefly to bring liquid drops to bottom of the tube.
- Incubate 15-20 minutes at room temperature.
- Add 200 μl DNA-Fect293-DNA complex drop-wise into each well containing cells and medium. Mix gently by rocking the plate back and forth.
- Incubate at 37^oC and 5% CO₂ in a humidified atmosphere.
- Transfection experiments are usually stopped after 24 to 48 hours and gene activity assessed. Cells growing in suspension are collected by centrifugation at 800 x g and then re-suspended in the desired medium or buffer.