

NEUROMICS



E18 Primary Rat Hippocampal Cells

Catalog Number: PC35101

Components : • **Approximately 1 x 10⁶ cells** (2 mls) E18 Sprague/Dawley or Fischer 344 hippocampal neurons.
• **12 mls Culture Media-** Neurobasal/B27/0.5 mM glutamine/25 µM glutamate culture medium **5 mls Hibernate** Shipping Media

Description: Primary Rat Hippocampal Cells are live neurons isolated from micro-surgically dissected regions of day 18 embryonic Sprague/Dawley or Fischer 344 rat brain. These cells are prepared fresh each week and shipped in a nutrient rich medium that keeps the cells alive for up to 7 days under refrigeration. The cells are ideal for a wide variety of applications including: transfection, pharmacology, electrophysiology, immunocytochemistry, and neuronal development studies.

Shipping/Storage: Primary Rat Hippocampal Cells are shipped refrigerated. Cells are stable for up to 7 days when stored at 4-8°C. It is recommended to plate the cells as soon as possible after receiving cells according to Application Notes outlined below.

References:

- [Scott E. Counts and Elliott J. Mufson. Noradrenaline activation of neurotrophic pathways protects against neuronal amyloid toxicity. Journal of Neurochemistry. Volume 113 Issue 3, Pages 649 - 660. Published Online: 28 Jan 2010.](#)
- [Karunya K. Kandimalla¹, Olenych G. Scott, Smita Fulzele¹, Michael W. Davidson, Joseph F. Poduslo. Mechanism of Neuronal versus Endothelial Cell Uptake of Alzheimer's Disease Amyloid β Protein. PLoS ONE 4\(2\): e4627. doi:10.1371/journal.pone.0004627.](#)
- [Jennifer M Laing and Laure Aurelian. ΔRR vaccination protects from KA-induced seizures and neuronal loss through ICP 10PK-mediated modulation of the neuronal-microglial axis. Genetic Vaccines and Therapy 2008, 6:1doi:10.1186/1479-0556-6-1.](#)

Application Notes

Materials Needed Not Provided

- Poly-D-lysine (Sigma P6407) for substrate
- Papain (Sigma P4762; or Worthington) for enzymatic dissociation
- Trypan blue to count cells to get proper plating density
- Sterile pipette tips or sterile Pasteur pipette
- Sterile centrifuge tubes
- Centrifuge to operate at 200xg
- Water bath at 30°C
- General cell culture supplies (culture plates, coverslips, etc.)
- Additional media
 - Neuromics' [NbActiv1-M36109](#)
 - OR
 - Neurobasal/B27 **without glutamate** ○ Glutamine (Invitrogen 35050-061)

Substrate Preparation

1. Prepare culture plate by coating with poly-D-lysine (0.15 ml/cm², 50 µg/ml, 135 kD) 1-20 hr., and rinse one time with 18 Mohm deionized water, and let dry.

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Preparation of Isolated Neurons

1. Store tissue at 4C until ready to use.
2. When ready to plate, make up 2 mls of enzymatic solution in shipping media without B27 (Hibernate-Ca; 5 mls supplied) containing 4 mgs (2mgs/ml) of papain. Make sure to sterile filter solution with a 0.2 micron filter after adding papain if source of enzyme is not sterile
3. Transfer the 2 ml of medium from the tissue tube into a 15 ml screw cap sterile tube; be careful not to disturb or remove tissue from the original tube. Save, do not discard.
4. Add 2 ml of media made in step 2 (Hibernate-Ca containing 2mgs/ml of papain). Incubate for 30 minutes at 30°C.
5. Remove enzymatic solution, again careful not to disturb or remove tissue, add back 1 ml of media saved in 15 ml tube.
6. Using a 1 ml pipettor with a sterile blue plastic tip, or a silanized 9-inch Pasteur pipette with the tip barely fire polished (preferable), suck the tissue with the medium into the pipette and immediately dispense the contents back into the same container. Take care not to create bubbles. Repeat this titration step about 10 times or until most all the cells are dispersed.
7. Let undispersed pieces settle by gravity for 1 min.
8. Transfer the dispersed cells (supernatant) into the 15 ml tube that contains the 1ml of media from Step 2, and gently mix the cells by swirling.
9. Spin the cells at 1,100 rpm (200xg) for 1 minute. Discard the supernatant while being careful not to remove any of the cells from the cell pellet.
10. Flick the tube a few times to loosen the cell pellet. Resuspend pellet in 1 ml of the provided B27/Neurobasal/0.5 mM glutamine medium. Resuspend cells by gently pipetting up and down. For E18 Hippocampus, media includes 25uM glutamate.
11. Aliquot 20 μ l and mix with 20 μ l of 0.4% trypan blue.
12. Count cells with a hemocytometer.
13. Further dilute the cells with B27/Neurobasal/0.5 mM glutamine to the desired plating density. We recommend 32×10^3 cells/2 cm^2 of substrate in 0.4 ml/2 cm^2 substrate.
14. Incubate the cells at 37°C with 5% CO₂ and/or 9% or 20% oxygen.
15. After 4 days or longer, neurons are well differentiated. If further culture is desired, change half of medium with fresh, warm NBActiv4 or B27/Neurobasal/0.5 mM glutamine, without glutamate. Change half the medium every 3-4 days. Additional media and media supplements will need to be purchased to culture neurons past 4-6 days.

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Customer Data

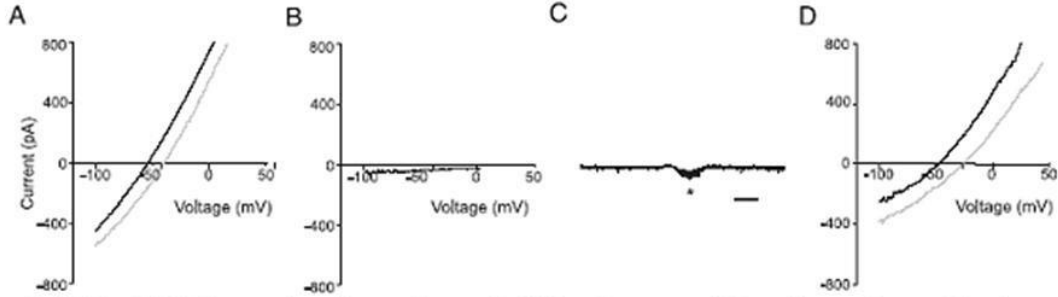
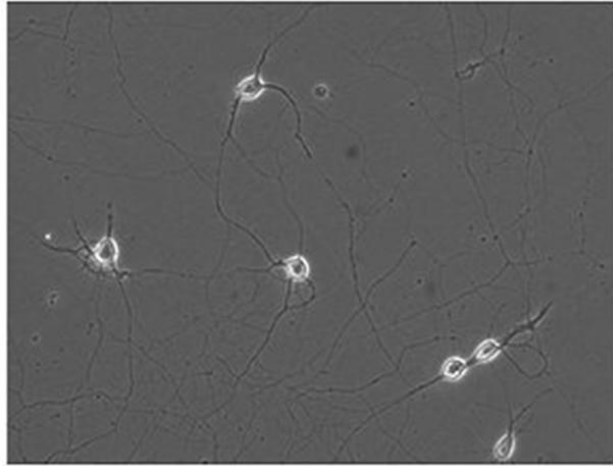


FIG. 9. NO-induced shift in E_{Cl^-} occurs in rat hippocampal neurons. **A:** GABA-gated currents recorded from a hippocampal neuron during voltage ramps before (black trace) and after (gray trace) NO application. (ruptured-patch, normal hippocampal external and internal solutions). **B–D:** current records elicited by the voltage protocol depicted in Fig. 7B. **B:** GABA-gated current collected in 0 external Cl^- . **C:** NO-dependent current (asterisk) recorded in 0 chloride indicates timing of NO arrival. Scale bar, 2 s. **D:** after returning to normal (TEA-H) external solution, a shift in E_{GABA} is revealed. The gray trace is recorded 30 s after return to normal $[Cl^-]_o$, and the black trace is recorded 1 min after return to normal $[Cl^-]_o$. Recordings in **A–D** are from the same cell. Recordings were made in the ruptured-patch configuration with TEA-H or 0 Cl^- -H external solutions and Cs^+ -H internal solution.

Hippocampal Neurons-1 Week in Culture. Courtesy of Emily McMains (Gleason Lab), LSU



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